Original Article

Angiotensin II is related to the acute aortic dissection complicated with lung injury through mediating the release of MMP9 from macrophages

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Abstract: Background: Acute aortic dissection (AAD) patients usually show concurrent lung injury mainly featured by hypoxemia. To date, no effective treatment method has been established for the AAD complicated with acute lung injury (ALI). Matrix metalloproteinases (MMPs), especially MMP2 and MMP9, have been considered to be closely related to the onset of aortic disease including AAD. To investigate the roles of MMP in the pathogenesis of AAD complicated with ALI, we determined the expression of MMP2 and MMP9 in serum and lung tissues of AAD patients. In addition, a new rat model of AAD complicated with ALI was established to investigate the pathogenesis of such complicated conditions. Methods and results: Angiotensin II (Ang II) and MMP9 were up-regulated in the AAD complicated with ALI patients compared to those of the AAD without ALI patients, normal individuals and the patients with non-ruptured aneurysm. Besides, massive macrophages with MMP9 expression was noticed in the lung tissues in the AAD complicated with ALI patients. On this basis, AAD complicated with ALI rat model was established based on BAPN feeding and infusion of Ang II. Obvious lung injury was observed in the BAPN+Ang II group compared to that of the BAPN group, together with macrophage accumulation in lung tissues, as well as over-expression of MMP9 in lung tissues. After interference of MMP antagonist, a large number of macrophages were still accumulated in the lung tissues, but the lung injury was obviously attenuated. After the interference of AT1 receptor, the number of macrophages in the lung tissues was obviously decreased and the lung injury was obviously relieved. Conclusions: Ang II is closely related to the lung injury at the early stage of AAD through mediating the release of MMP9 in the macrophages in the lung tissues.

Keywords: Acute aortic dissection, animal model, lung injury, MMP9

Introduction

Acute aortic dissection (AAD), the most frequent and catastrophic manifestation of acute aortic syndrome, causes great threats to the public health worldwide [1-3]. Despite the recent progression of diagnosis and treatment, the mortality rate of AAD is still high. In clinical practice, AAD is frequently accompanied by acute respiratory failure such as acute respiratory distress syndrome (ARDS) and/or acute lung injury (ALI). Besides, concurrent ALI before surgery is an independent risk factor for post-operative ARDS [6-8]. Several factors have been considered as the common causes for ALI, including hypotension, pulmonary infection, as well as radiation injury and mechanical injury in lung [9-11]. However, many AAD patients show concurrent ALI despite the absence of such factors. Therefore, it is reasonable to speculate that there might be a close relation between AAD and ALI. Respiratory failure in AAD patients appears to be closely correlated with the aortic injury and even the prognosis. However, the exact mechanism is far from clear. Thus, the unknown pathogenesis of AAD complicated with ALI deserves a study of establishing animal model for the clinical and experimental research.

Currently, no AAD complicated with ALI model has been established as most of the studies have focused on the induction of AAD model. According to the previous description, the induction of AAD animal models is mainly depend on the surgical methods [4, 5] or the β-aminopropionitrile (BAPN) treatment. However, all these animal models are restricted to aortic dissection, and the incidence of acute lung injury (ALI) was extremely lower. In this


AngII mediated release of MMP9 from macrophages involved in the AAD merged ALI

study, a new AAD complicated ALI model was established based on the feeding of BAPN and the infusion of Ang II. On this basis, we aim to investigate the exact pathogenesis of AAD complicated with ALI in rats. Our study reveals Ang II is closely related to the lung injury at the early stage of AAD through mediating the release of MMP9 in the macrophages in the lung tissues.

Materials and methods

Human blood sample

Fifty-eight newly diagnosed AAD patients admitted in the ICU of our hospital from September 2014 to July 2015 were included in this study. Twelve matched individuals and 12 non-ruptured aortic aneurysm patients were registered. The diagnosis of AAD and aortic aneurysm was performed based on the CT scan and ultrasonic examination. Patients admitted to our hospital 7 days after the onset of disease were excluded from the study. Those with cancer, chest trauma and pulmonary infection within one month were excluded from this study. Blood sample was collected and PaO2/FiO2 was determined 1 h after admission. Twenty-one AAD patients showed combined hypoxemia before the surgery. The patient characteristics were listed in Table 1. Written informed consent was obtained from each patient. The study protocols were approved by the Ethical Committee of the Renmin Hospital of Wuhan University.

Upon collection of lung tissues, the samples were fixed and embedded. Afterwards, H&E staining and immunohistostaining were performed according to the previous description [12]. The images were observed using a CKX41SF light microscope (Olypus Corporation, Tokyo, Japan) to determine the histopathological changes in lung tissues, as well as the expression of MMP9 and CD68.

Development of AAD complicated lung injury model in rats

Patients with AAD complicated with lung injury showed elevation of serum Ang II. On this basis, AAD complicated with lung injury rat model was established using BAPN treatment combined with pumping of Ang II. The male SD rats (3-week-old) were provided by the Animal Center of Wuhan University and were divided into three groups: control group, wild type male SD rats fed by normal diet for 4 weeks; BAPN group, wild type male SD rats fed by normal diet for 4 weeks, combined with treatment of BAPN (0.1 g/kg/d, Sigma-Aldrich, St. Louis, MO) for 4 weeks; BAPN+Ang II group, wild type male SD rats fed by normal diet, combined with treatment of BAPN (0.1 g/kg/d) for 4 weeks, and pumping of Ang II (1 μg/kg/min, Sigma-Aldrich, St. Louis, MO) for 24 hours. Only the animals with formation of aortic dissection were used for the subsequent study. All animal handling was performed in accordance with the Wuhan Directive for Animal Research and the current Guidelines for the Care and Use of Laboratory Animals published by the National Institutes of Health (NIH publication no. 85-23, revised 1996). Rats were sacrificed after successfully anesthetized with 2% phenobarbital (40 mg/Kg). Extensive measures were taken to lower the sufferings of the animals. The animal study

<table>
<thead>
<tr>
<th>Table 1. Clinical features of the patients</th>
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<tr>
<td>Control (n=12)</td>
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<tr>
<td>----------------</td>
</tr>
<tr>
<td>Age, yr</td>
</tr>
<tr>
<td>Male sex, n %</td>
</tr>
<tr>
<td>Average duration from onset, h</td>
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<tr>
<td>Smoking, n (%)</td>
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Histopathological examination

Pulmonary tissues of aortic dissection were obtained from 2 cadavers with AAD combined with ALI received no surgery before. Pulmonary tissues obtained from the organ donors (n=2) served as the control. The study protocols were approved by the Ethical Committee of the Renmin Hospital of Wuhan University.

ELISA

Serum matrix metalloproteinase 2 (MMP2), MMP9, and Ang II were measured using commercial ELISA kits (category No. EK0657, EK0465 and EK0459, Biofavor Biotech Service Co., Ltd. Wuhan, China) according to the manufacturer’s instructions. All tests were carried out at least in triplicate.
AngII mediated release of MMP9 from macrophages involved in the AAD merged ALI

was approved by the Ethical Committee of the Renmin Hospital of Wuhan University.

Verification of AAD complicated lung injury model in rats

To confirm whether the Ang II involves in the recruitment of macrophages in this study, we determined the accumulation of macrophages in the lung tissues after interference of valsartan (10 mg/kg per day, Norvatis, Beijing, China). For pharmacological depletion of MMP9, BAPN-fed rats were administered by gastric lavage with a broad-spectrum MMP inhibitor, ONO-4817 (300 mg/kg per day, Bludgebio Co., Ltd, Shanghai, China, category No. 2628), daily for 2 days before Ang II administration until sacrifice.

Reverse transcription PCR

Total RNA was extracted from the lung tissues using Trizol reagent (Invitrogen, Shanghai, China) according to the manufacturer's instructions. Reverse transcription PCR was carried out using the specific primers targeting MMP9 gene as previously described [13]. PCR primers were synthesized by the Biofavor Biotech Service Co., Ltd. (Wuhan, China). GAPDH served as internal standard. The primer sequences for MMP9 were as follows: forward, 5'-CAGTGCCAGCTCGTCTCAT-3'; reverse, 5'-AGGGGCCATCCAGCTTCTC-3'. GAPDH were as follows: forward, 5'-CAGTGCCAGCTCGTCTCAT-3'; reverse, 5'-AGGGGCCATCCAGCTTCTC-3'.

Immunohistological examination

Sections (4 μm thick) were cut from the paraffin-embedded lung specimens obtained from the cadavers. MMP9 was labeled using rat monoclonal antibody purchased from Proteintech (category No. 10375-2-AP, Wuhan, China). The CD68 of macrophages were labeled using the nonoclonal antibody purchased from Biolegend (category No. 333801, San Diego, CA, China). MMP9 and CD68 were labeled with Cy3 (category No. BA1032, Boster Co., Ltd. Wuhan, China) and fluorescein isothiocyanate (FITC, category No. BA11.0, Boster Co., Ltd. Wuhan, China). The images were observed under the Eclipse 80i fluorescence microscope (Nikon, Tokyo, Japan). Immunohistochemistry image analysis was performed using IPP6.0 software.

Additional methods

For the additional methods, the expression of malondialdehyde (MDA) content and the weight-to-dry weight ratios (W/D ratios) and superoxide dismutase (SOD) in lung were determined. All the tests were carried out at least in triplicate.

Statistical analysis

All statistical analysis were performed with SPSS 20.0 for windows. The data were presented as mean ± standard deviation. Human blood sample data were analyzed with 2-sample t-tests, and the occurrence rate of rats AAD complicated lung injury was analyzed with Fisher's exact test. P value less than 0.05 was regarded as significant.

Results

Roles of MMP9 and Ang II in the onset of AAD plus ALI

AAD patients showed remarkable decrease of \( PO_2 \) and \( SpO_2 \); No statistical difference was
AngII mediated release of MMP9 from macrophages involved in the AAD merged ALI

noticed in the age and gender in the AAD patients, normal individuals and the non-ruptured aneurysm patients. Compared with the normal individuals and non-ruptured aneurysm patients, the levels of PO2 and SpO2 were remarkably decreased in the AAD patients (Figure 1). Among the 58 AAD patients, 21 (36.2%) showed hyoxemia with a PaO2/FiO2 of ≤ 300 mmHg. This revealed patients with AAD showed lung injury mainly featured by hyoxemia.

Elevation of MMP9 and Ang II in blood samples from AAD complicated with ALI patients: The concentration of Ang II and MMP9 in the patients with AAD complicated with ALI was remarkably elevated compared to the normal individuals, the non-ruptured aneurysm patients, and the AAD patients without ALI (Figure 2), which implied Ang II and MMP9 may be associated with the onset of AAD complicated with ALI. Whereas, the expression of MMP2 in the AAD complicated with ALI patients showed no difference compared with the other groups.

MMP9 in lung tissues from AAD complicated with ALI patients was mainly released from macrophage: Abundant MMP9 positive cells were located mainly in the lung tissues in the AAD complicated with ALI patients as revealed by immunolocalization, while rare expression of MMP9 was identified in the normal tissues. In addition, a large number of neutrophils and macrophages were noticed in the lung tissues from the AAD complicated with ALI patients. This indicated the MMP9 in the lung tissues of AAD complicated with ALI patients may derive from the neutrophils or macrophages. To confirm this, double immunofluorescence labeling was performed to the MMP9/MPO (neutrophils) and MMP9/CD68 (macrophages), and the results revealed high colocalization of MMP9 and CD68 in the cells, while rare colocalization of MMP9 and MPO was observed in the cells (Figure 3).

Taken together, we concluded that AAD complicated with ALI patients showed remarkable elevation of MMP9 and Ang II. MMP9 was mainly released from the macrophages. Ang II may
AngII mediated release of MMP9 from macrophages involved in the AAD merged ALI

Figure 3. MMP9 expression and neutrophils and macrophages in the lung tissues of cadavers with AAD complicated with lung injury using immunofluorescence labeling. Colocalization of MMP9 and CD68 was observed in the lung tissues in AAD patients, but rare colocalization of MMP9 and MPO was observed, which indicated the MMP9 was mainly derived from the macrophages in lung tissues. Green color: MPO (neutrophil) and CD68 (monocyte/macrophage), red color: MMP9, blue color: nucleus.
AngII mediated release of MMP9 from macrophages involved in the AAD merged ALI

AngII, one of the representative vasopressors, is considered to be closely related to in the pathogenesis of several vascular diseases including AAD. Also, it is known to induce MMP9 expression in macrophage [14, 15]. Besides, it involves in inflammation through recruiting inflammatory cells in local tissues [16]. On this basis, we hypothesize that AngII may modulate the release of MMP9 from macrophage and involve in recruitment of macrophages in lung tissues. In this study, animal model was used to investigate the roles of AngII in the modulation of macrophage in the subsequent section.

Confirmation of AngII roles in animal models

AngII infusion contributes to the establishing of AAD complicated with ALI rat model: Based on the remarkable elevation of AngII and MMP9 in AAD complicated with ALI patients, the rat model was established through feeding of BAPN and subcutaneous infusion of AngII. Compared with the rats received BAPN feeding, a higher incidence of lung injury was observed in the rats subject to BAPN feeding and infusion of AngII (85.7% vs. 28.6%). Procedures were performed to verify whether the rat model was successfully induced. For the HE staining, massive infiltration of inflammatory cells were noticed in the lung tissues in the BAPN+AngII group, together with injury of pulmonary alveoli. Besides, red blood cells, inflammatory cells and exudate were noticed in the alveolar space (Figure 4).

Further, the malondialdehyde (MDA) content and the weight-to-dry weight ratios (W/D ratios) in lung was remarkably elevated together with obvious decrease of superoxide dismutase (SOD) activities (Table 2). Taken together, it is reasonable to conclude that obvious lung injury was observed in the BAPN+AngII group. On this basis, AngII may be related to the establishing of AAD complicated with ALI rat model.

Table 2. Determination of MDA, SOD and W/D in each group

<table>
<thead>
<tr>
<th>Group</th>
<th>MDA (nmol/mgprot)</th>
<th>SOD (U/mgprot)</th>
<th>W/D</th>
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<tbody>
<tr>
<td>Normal control</td>
<td>1.34±0.31</td>
<td>48.19±4.24</td>
<td>3.25±0.37</td>
</tr>
<tr>
<td>BAPN group</td>
<td>1.77±0.26</td>
<td>41.74±2.83</td>
<td>3.48±0.54</td>
</tr>
<tr>
<td>BAPN+AngII group</td>
<td>4.84±0.49*</td>
<td>20.70±3.17*</td>
<td>4.57±0.66*</td>
</tr>
</tbody>
</table>

*p<0.05, compared with normal control.

Up-regulation of MMP9 expression in the rats model with AAD complicated with ALI: MMP9 expression was determined using reverse transcription polymerase chain reaction in the lung tissues of rats 24 hours after AngII infusion. The expression of MMP9 mRNA was remarkably elevated in the BAPN+AngII group (Figure 5A) compared to that of the BAPN group and control group, respectively. Immunohistochemistry results indicated massive oedema in the lung tissues, infiltration of inflammatory cells in alveolar mesenchyma, as well as obvious elevation of MMP9 in the BAPN+AngII group (Figure 5B).

To investigate the roles of MMP9 in the pathogenesis of AAD complicated with ALI, ONO-4817 (MMP antagonist) was administrated in the BAPN+AngII group. The incidence of AAD was reduced by 28.6%. About 20.0% of the...
AngII mediated release of MMP9 from macrophages involved in the AAD merged ALI

AAD rats showed ALI. The lung injury was remarkably attenuated in AAD rats (Figure 6). Compared with the BAPN+Ang II groups, the MDA and W/D ratio were remarkably decreased in the group after interference of ONO-4187 (P<0.05). Besides, remarkable elevation was noticed in the SOD level in the group with interference of ONO-4187 compared with that of the BAPN+Ang II group (Table 3, P<0.05).

Ang II induced macrophages recruitment to lung injury through AT1 receptor independent from MMP9 expression: Immunohistological analysis of BAPN+Ang II group indicated adhesion and infiltration of macrophages in the lung tissues combined with obvious up-regulation of MMP9. On this basis, it is reasonable to conclude that Ang II could induce recruitment of macrophages. However, such fact could not exclude the following possibility: Ang II may induce the release of MMP9 in the lung tissues, while the latter could recruit macrophages and the consequent release of MMP9. To confirm whether the MMP9 involves in the recruitment of macrophages in this study, we determined the accumulation of macrophages in the lung tissues after interference of valsartan or ONO-4817, respectively. As shown in Figure 7, remarkable attenuation was noticed in the number of macrophages in the lung tissues subject to valsartan treatment, while no statistical difference was noticed in the macrophage count in the lung tissues subject to ONO-4817 interference. Taken together, Ang II induced the

Figure 5. MMP9 expression in the lung tissues in BAPN+Ang II group. A: MMP9 mRNA expression in each group using RT PCR. B: Immunohistochemistry analysis of MMP9 in each group: a: control; b: BAPN group; c: BAPN+Ang II group. The images were observed under a magnification of 400×.

Figure 6. HE staining of the lung tissues in normal group (A), BAPN+Ang II group (B) and ONO-4817 group (C). In the BAPN+Ang II group, obviously oedema was observed in the alveolar space and infiltration of inflammatory cells in the alveolar mesenchyma, as well as injury of alveoli. In the ONO-4817 group, the pulmonary injuries were obviously attenuated compared with that of the BAPN+Ang II group. The images were observed under a magnification of 200×.
Ang II mediated release of MMP9 from macrophages involved in the AAD merged ALI

**Table 3.** MDA, SOD and W/D changes after interference of ONO-4187

<table>
<thead>
<tr>
<th>Group</th>
<th>MDA (nmol/mgprot)</th>
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<td>20.70±3.17</td>
<td>4.57±0.66</td>
</tr>
<tr>
<td>ONO-4817 group</td>
<td>2.94±0.31*</td>
<td>34.74±2.83*</td>
<td>3.75±0.43*</td>
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</table>

*P<0.05, compared with BAPN+Ang II group.

Ang II has been acknowledged as an important hormone for the renin-angiotensin system (RAS). Ang II could bind the receptor types AT1R and AT2R. It plays a crucial role in the recruitment of inflammatory cells and induction of monocytes and macrophages. Moreover, it could stimulate the release of IL-1, IL-6, IL-8, tumor necrosis factor-α (TNF-α), and macrophage chemoattractant protein-1 (MCP-1), which finally results in ALI [20-22]. In a previous study, Kaliappan et al. revealed Ang II played a crucial role in the pathogenesis of aortic aneurysm through recruiting macrophages in the aortic wall [23]. In addition, monocytes are important target cells of Ang II and express both AT1 receptor and AT2 receptor. These cells have substantial roles in promoting inflammation, foam cell formation and MMP secretion [24, 25]. In our study, compared with the BAPN group, the cell aggregation of macrophages was more obvious in the BAPN+Ang II group, and such phenomenon was obvious attenuated after administration of AT1 antagonist. This indicated the induction of macrophages recruiting mediated by Ang II was depend on the AT1 receptor. Moreover, Ang II contributed to the expression of EMMPRIN and MMP9 in macrophages through protein kinase C and Rho kinase inflammatory signaling pathway. Such biological process was also depend on the AT1 receptor rather than the AT2 receptor [26, 27]. In a previous study, up-regulation of EMMPRIN could contribute to the autocrine and paracrine secretion of MMPs, which subsequently enhanced the migration of monocytes and macrophages into the lung mesenchyma [28]. Recently, extensive studies reveal Ang II play crucial roles in the pathogenesis of AAD [18, 29-32]. For example, Kurihara et al. reported Ang II mediated MMP9 release in the neutrophils in the aortic wall, and was considered as the major cause for the pathogenesis of AAD [18]. This implied Ang II contributed significantly to the onset of inflammation through modulating the recruiting of inflammatory cells in various tissues.

**Discussion**

The pathogenesis of AAD complicated lung injury is still unknown. Recent studies reveal the onset of the disease is closely related to the acute inflammatory reactions, C reactive protein, D2 dimer, body temperature and the scale of aortic dissection [17]. In this study, the level of Ang II showed remarkable increase in the patients with AAD complicated with lung injury compared to that of the normal individuals. In addition, in the lung tissues derived from the cadavers of AAD complicated with lung injury, a large number of MMP9 expressing macrophages were accumulated in the pulmonary mesenchymal. Taken together, it is reasonable to speculate that MMP9 and Ang II may involve in the onset of AAD complicated with lung injury.

Based on these results, we established an rat model of AAD complicated with lung injury based on the pumping of Ang II following the BAPN treatment. Up to now, BAPN is the most frequently drug for the induction of AAD animal models. It is a lysyl oxidase inhibitor, which could induce cystic medial degeneration in the aortic wall in rats. In the previous studies, BAPN could induce AAD in immature rats [18], however, no obvious lung injury was induced. In this study, AAD complicated with lung injury model was established using BAPN feeding combined with infusion of Ang II. Our study confirmed such treatment could induce the AAD complicated lung injury successfully.

In this study, the level of Ang II in the AAD complicated with lung injury was obviously higher than that of the normal control, the non-ruptured aneurysm group and AAD without ALI patients. Unlike the previous animal models based on BAPN [19], our study showed the lung injury was severe in the rat models subject to infusion of Ang II combined with BAPN feeding. Besides, the expression of MMP9 was higher in the lung tissues. This indicated that Ang II may play a crucial role in the onset of AAD complicated lung injury.
AngII mediated release of MMP9 from macrophages involved in the AAD merged ALI

ALI/ARDS is a complex condition of acute, progressive respiratory failure [33] caused by inflammatory reactions in the lung mediated by inflammatory cells especially the macrophages and the dysfunction of blood gas barrier induced by inflammatory disorder [34, 35]. MMP9 is a proteinase involved in the degradation and denaturation of type III-V collagen protein and elastin [36], which were the major components of the extracellular matrix and basilar membrane of blood vessels. Therefore, MMP9 could degrade the extracellular matrix and basilar membrane in the pulmonary capillary. In addition, it could destroy the integrity of basilar membrane, and contribute to the migration of inflammatory cells in the inflammation. Moreover, it could affect the alveolocapillary barrier and result in increase of capillary permeability, infiltration of inflammatory cells and formation of oedema in lung [37-42]. In this study, patients with AAD complicated with ALI showed massive accumulation of MMP9 expressing macrophages in lung tissues, and such phenomenon was obviously attenuated after interference of MMPs antagonist without affecting the accumulation of macrophages.

There are still limitations for our study. Firstly, the animal based model could not mimic the pathogenesis of AAD complicated with ALI in human completely. Secondly, we can not exclude the involvement of other inflammatory cells in the induction of ALI.

In conclusion, AAD complicated with ALI rat model was established based on BAPN feeding and infusion of Ang II. Ang II and MMP9 are considered as the potential diagnostic indices of AAD complicated with ALI. Early-stage administration of AT1R blocking agent and MMPs antagonist may prevent or attenuate the pathogenesis and development of ALI. Besides, Ang II is closely related to the lung injury at the early stage of AAD through mediating the release of MMP9 in the macrophages in the lung tissues.

Disclosure of conflict of interest

None.

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AngII mediated release of MMP9 from macrophages involved in the AAD merged ALI

References

AngII mediated release of MMP9 from macrophages involved in the AAD merged ALI


