MicroRNA-495 suppresses human renal cell carcinoma malignancy by targeting SATB1

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Abstract: Deregulated expression of miRNAs is related to progression and initiation of human cancers. Although miR-495 has identified in various tumors, its expression and function in renal cell carcinoma (RCC) is still unknown. In this study, we found that the expression of miR-495 was downregulated in RCC cell lines and tissues. Ectopic expression of miR-495 induced G0/G1 phase arrest and suppressed cell proliferation and migration in RCC cell lines. We further validated SATB1 was a direct target of miR-495 in RCC. In addition, re-expression of SATB1 reversed the miR-495-induced inhibition of cell proliferation and migration. These data suggest that miR-495 functions as a tumor suppressor and may be a promising therapeutic target in RCC in the future.

Keywords: Renal cell carcinoma, microRNAs, miR-495, SATB1

Introduction

Renal cell carcinoma (RCC) is the most common neoplasm of the adult kidney, which accounts for approximately 85% of all primary malignant kidney tumors as well as 3% of cancers in adults [1-4]. Surgery is often curative for localized disease; however, a lot of these patients develop metastatic or relapses diseases, which are associated with poor prognosis [5-8]. The 5-year-survival rate of patients who diagnosed at the metastatic stage was only 9% [9-11]. Therefore, there is a crucial need to found new biomarkers and targeted therapies for this aggressive malignancy.

MicroRNAs (miRNAs), approximately 19-24 nucleotides in length, are highly conserved regulatory molecules that modulate gene expression through imperfect complementary sequence pairing to the 3’ untranslated region (3’UTR) of target genes resulting in either mRNA degradation or translational repression [12-14]. Several studies have reported that miRNAs play key roles in developmental regulation, cell proliferation, differentiation, invasion and apoptosis [15-17]. Abnormal expressions of miRNAs have been found in several types of human cancers, including cervical cancer, bladder cancer, breast cancer, renal cell carcinoma, hepatocellular and ovarian cancers [12, 18-23]. They may play important roles in the progression and development of cancers similar to those played by tumor suppressor genes or oncogenes [24-27].

In this study, we demonstrated that miR-495 expression level was frequently downregulated in human RCC cell lines and tissues and overexpression of miR-495 suppressed RCC cell proliferation and migration. Moreover, SATB1 was identified as a direct target of miR-495 and re-expression of SATB1 reversed the miR-495-induced inhibition of cell proliferation and migration.

Materials and methods

Ethics statement

Patients gave written informed consent. This study was approved by the ethical board of Renmin Hospital of Wuhan University and complied with the Declaration of Helsinki.

Tissues and cell lines

RCC tissues and adjacent normal tissues were collected from patients undergoing surgery at
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Figure 1. miR-495 was downregulated in RCC cell lines and tissues (A) qRT-PCR analysis was used to detect the expression of miR-495 in four human renal cell carcinoma cell lines (769-P, 786-O, A498, SN12-PM6) and one normal renal cell line (HK-2). (B) qRT-PCR analysis was used to detect the expression of miR-495 in clinical RCC specimens and normal tissues. (C) The relative expression of miR-495 was downregulated in clinical RCC specimens compared with their corresponding nontumor tissues.

Figure 2. Re-expression of miR-495 suppressed RCC cell proliferation (A) qRT-PCR was performed to measure the miR-495 expression in 786-O cells at 48 hours after miR-495 mimic transfection. (B) Cell cycle assays showed that 786-O cells transfected with miR-495 mimics had an obvious cell cycle arrest at the G0/G1 phase. (C) Re-expression of miR-495 suppressed 786-O cells proliferation using CCK-8 assay.

Figure 3. Overexpression of miR-495 inhibited RCC cell migration. Overexpression of miR-495 repressed the 786-O cells migration.

our hospital. The samples were immediately snap-frozen in liquid nitrogen until protein or RNA extraction. The human RCC cell lines, 769-P, 786-O, A498, SN12-PM6 and one normal renal cell line (HK-2) were purchased from the Cell Bank of the Chinese Academy of Sciences.
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These cells were maintained in Dulbecco’s modified Eagle’s medium (Invitrogen) supplemented with 20% FBS.

**Transient transfection**

MiR-495 mimics and scramble were purchased from GenePharma (GenePharma, China). MiR-495 mimics or scramble was transfected into cells by using Lipofectamine 2000 (Invitrogen, Canada) according to the manufacturer’s instructions.

**Cell proliferation, cell cycle and colony formation**

CCK-8 assay was performed to evaluate the cell proliferation according the manufacturer’s instructions.

Optical density (OD) was measured at 450 nm an enzyme immunoassay instrument (BioRad, Hercules, CA, USA). For cell cycle experiment, cells were fixed in 95% ethanol, incubated at -20°C overnight and resuspended in FACS solution. Cells were detected by using a FACS Calibur flow cytometer (BD Biosciences). For colony formation assay, cells were cultured for 7 days. Then cells were then fixed with 4% formaldehyde and stained with 1.0% crystal violet.

**Real-time RT-PCR**

Total RNA was extracted using TRIzol reagent (Invitrogen). SYBR-Green PCR master mix (Applied Biosystems, Inc. Foster City, CA, and USA) was performed on the 7500 Real-time PCR System (Applied Biosystems). The expression of U6 or GAPDH was used as control. The primer sequences were: SATB1 forward: 5’-GAGGAAGGCTTGGGAGTA-3’, reverse: 5’-GGGCAGCAGCTATGTG-3; GAPDH forward: 5’-GGGAGGCAACGGATTTGGTCGTAT-3’ and reverse: 5’-AGCCTTCTCCATGGTGAGTGAAC-3’.

**Dual-luciferase reporter assay**

Luciferase assay was performed as previously reported [28]. SATB1 3’-UTR containing the putative miR-495 binding site or a mutant sequence was designed and inserted into the downstream of the firefly luciferase reporter.
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(Promega, Madison, WI, USA). The cultures transiently cotransfected with miR-495 scramble and vector (contain either WT or MUT 3’-UTR). Luciferase activity was performed by Dual Luciferase Reporter Assay System.

Western blotting

Western blotting was performed as previously described [29]. Protein was separated by electrophoresis, transferred to membranes and were probed with specific primary antibody. The membranes were incubated with the secondary antibody conjugated to horseradish peroxidase (HRP). Protein was visualized by chemiluminescence using the ECL detection system (BeyoECL Plus, Beyotime). Anti-SATB1 and GAPDH antibodies were purchased from Abcam (Cambridge, MA, USA).

Migration assay

Cell migration was performed using a wound-healing assay. The cell monolayer was scraped using a P-20 micropipette tip. The initial gap length and the residual gap length after wounding were measured by photomicrographs.

Statistical analysis

All experiments were performed in triplicate and analyzed for significant differences between two groups were analyzed using Student’s t test; One-way ANOVA was performed to analyze the more than two groups. P < 0.05 was considered statistically significant. The data are showed as means ± SD.

Results

miR-495 was downregulated in RCC cell lines and tissues

The expression of miR-495 was lower in all four human renal cell carcinoma cells lines (769-P, 786-O, A498, SN12-PM6) than in one normal renal cell line (HK-2) (Figure 1A). Moreover, qRT-PCR data demonstrated that the expression level of miR-495 was also significantly

Figure 5. Re-expression of SATB1 reversed the miR-495-induced inhibition of cell proliferation and migration. A. The protein expression of SATB1 was measured by using Western blot. B. The data of CCK8 assay showed that re-expression of SATB1 increased the miR-495 overexpressing 786-O cells proliferation. C. Cell cycle assays showed that miR-495 overexpressing 786-O cells transfected with SATB1 vector had an obvious cell cycle at the S phase. D. The migration abilities of miR-495 overexpressing 786-O cells were increased after SATB1 transfection.
lower in clinical RCC specimens and compared with normal tissues (Figure 1B and 1C).

Re-expression of miR-495 suppressed RCC cell proliferation

We aimed to address the phenotypes of 786-O cells stably expressing miR-495 (Figure 2A). Cell cycle assays showed that 786-O cells transfected with miR-495 mimics had an obvious cell cycle arrest at the G0/G1 phase (Figure 2B). Re-expression of miR-495 suppressed 786-0 cells proliferation using CCK-8 assay (Figure 2C).

Overexpression of miR-495 inhibited RCC cell migration

Migration assays were performed to measure the function of miR-495 in cell migration. As shown in Figure 3, cell migration was significantly repressed in the group transfected with miR-495 compared with those in the scramble group.

miR-495 downregulated SATB1 expression by directly targeting its 3’-UTR

The data of TargetScan showed SATB1 has the putative target of miR-495 (Figure 4A). Luciferase reporter assay showed that the luciferase reporter activity decreased approximately 67% in the 786-O cells containing the SATB1 WT 3’UTR fragment (Figure 4B). Moreover, we found that the ectopic expression of miR-495 suppressed the SATB1 mRNA and protein level in 786-O cells by using qRT-PCR and Western blot (Figure 4C and 4D).

Re-expression of SATB1 reversed the miR-495-induced inhibition of cell proliferation and migration

Rescue experiment to confirm that miR-495 acts as a tumor suppressor in renal cell carcinoma cells by regulating SATB1 expression. The expression of SATB1 was overexpressed by using SATB1 vector (Figure 5A). We rescued the expression of SATB1 in miR-495 overexpressing 786-O cells. CCK8 assay showed that re-expression of SATB1 increased the miR-495 overexpressing 786-0 cells proliferation (Figure 5B). Cell cycle assays showed that miR-495 overexpressing 786-0 cells transfected with SATB1 vector had an obvious cell cycle at the S phase (Figure 5C). Furthermore, the migration abilities of miR-495 overexpressing 786-O cells were increased after SATB1 transfection (Figure 5D).

Discussion

RCC is the third most common urological cancer and is one of the most common types of cancers [3, 30, 31]. Despite the great improvement in cancer therapy, metastatic RCC presents a 5-year survival rate of 0-10% [32-34]. Therefore, it is important to search new treatment strategies. In this study, we demonstrated that miR-495 expression level was down-regulated in human RCC cell lines and tissues and overexpression of miR-495 suppressed RCC cell proliferation and migration. Moreover, SATB1 was identified as a direct target of miR-495 and re-expression of SATB1 reversed the miR-495-induced inhibition of cell proliferation and migration. Taken together, our study suggests that miR-495 acts a tumor suppressor gene in RCC.

MiR-495 was reported to act as a tumor suppressor gene or an oncogene in a lot of cancers including non-small cell lung cancer, breast cancer, glioblastoma, gastric cancer and leukemia [35-38]. For example, miR-495 was down-regulated in gastric carcinoma samples and overexpression of miR-495 inhibited gastric cancer cell migration and invasion by targeting the PRL-3 oncogene [35]. Another study showed that miR-495 likely functions as a tumor suppressor in acute myeloid leukemia (AML) with mixed lineage leukemia (MLL) rearrangements by targeting essential leukemia-related genes [37]. However, it has been demonstrated that miR-495 acts as an oncogene in breast cancer via downregulation of E-cadherin and REDD1 [38]. Cao et al. also showed that miR-495 could facilitate breast cancer progression through the repression of JAM-A [39]. In our study, our data showed that the expression of miR-495 was lower in four human renal cell carcinoma cells lines (769-P, 786-O, A498, SN12-PM6) than in one normal renal cell line (HK-2). The expression level of miR-495 was also significantly lower in clinical RCC specimens and compared with normal tissues. Moreover, re-expression of miR-495 had an obvious cell cycle arrest at the G0/G1 phase and suppressed RCC cells proliferation and invasion. These results confirmed that miR-495 acts as a tumor suppressor gene in RCC.
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It is crucial to explore the molecular mechanism underlying miR-495 function in RCC. The putative target of SATB1 for miR-495 was detected by the TargetScan program. Luciferase reporter assay showed that the luciferase reporter activity decreased approximately 67% in the RCC cells containing the SATB1 WT 3'UTR fragment. To test this assumption, we detected the mRNA and protein expression of SATB1 after miR-495 overexpression. We found that ectopic expression of miR-495 suppressed the SATB1 mRNA and protein level in RCC cells. Furthermore, re-expression of SATB1 reversed the miR-495-induced inhibition of cell proliferation and migration. Our results suggest that SATB1 is a functional downstream target of miR-495 in RCC. Previous study showed that the levels of SATB1 mRNA and protein were increased in human RCC tissues and SATB1 knockdown inhibited the proliferation, migration and invasion of 786-O cells [40]. However, the detail mechanism of downregulation of SATB1 is still unknown. Our data showed that the ability of miR-495 to repress SATB1 expression may provide one such mechanism of post-transcriptional control of SATB1.

In conclusion, this study provided novel evidence that miR-495 functions as a tumor suppressor miRNA in RCC through inhibiting SATB1 expression. Our data suggested that this miRNA could be a potential target for the treatment of RCC in future.

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Disclosure of conflict of interest

None.

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